

Multiple Antibiotic Resistance Patterns of the *Enterobacteriaceae* in the Untreated Municipal Sewage

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Dear Editor,

The increasing incidence of bacterial infection and gradual rise in resistance in *Enterobacteriaceae* against the available antibiotics, has highlighted the need to find more alternative antibacterial agents from other source [1]. Despite the pervasive incidence in many parts of the world of this family, surprisingly diminutive is recognized concerning the occurrence of such bacteria in raw sewage in India, although the work was done on hospital sewage [2]. The pathogen residing in the sewage wastes can migrate to ground and surface water and can contaminate them as well, in addition the increased and widespread use of antibiotics by human and animal have resulted in the increased antibiotic resistance in bacteria which can create havoc and can have a devastated impact on public health. The study was undertaken to assess the multiple antibiotic resistance pattern of the *Enterobacteriaceae* in the untreated municipal sewage.

An unprocessed sewage sample having 100ml volume was collected from municipal Wastewater Treatment Plant (WTP) located in Pholriwari village, Jalandhar city having capacity, 100 metric liters per day, from September 2014 to December 2014, every month, by composite sampling method described by Central Pollution Control Board (CPCB), India, in a sterile glass bottle [3]. Sewage was serially diluted then 0.1ml volume was spread onto MacConkey agar plate and incubated at 37°C for 24hours. Total 31 isolates belonging to 16 species of the genera *Escherichia*, *Providencia*, *Proteus*, *Serratia*, *Yersinia*, *Kluyvera*, *Budivicia*, *Salmonella*, *Shigella* and *Hafnia* were identified after biochemical screening using KB001 HiMMViC Biochemical test Kit by Himedia (indole, methyl red, Voges Proskauer's, citrate utilization, glucose, adonitol, arabinose, lactose, sorbitol, mannitol, rhamnose and sucrose). These were tested against 11 different antibiotics, namely amikacin (30µg), ampicillin (10µg), cefixime (5µg), ceftazidime (30µg), chloramphenicol (30µg), ciprofloxacin (5µg), co-trimoxazole (25µg), doxycycline hydrochloride (30µg), gentamycin (10µg), nalidixic acid (30µg) and tetracycline (30µg) by disc diffusion method [4]. The results were interpreted according to Clinical and Laboratory Standards Institute (CLSI) guidelines (2012) and Multiple Antibiotic Resistance (MAR) index of each individual isolate was also taken into account by method described by Krumperman 1983 [5,6]. All the isolates were resistant to at least two of the antibiotics, as shown in [Table/Fig-1], and the following resistant patterns of all the isolates against 11 antibiotics were found in percentage: Amp¹⁰ (93.54) > Caz³⁰ (74.19) = Cfm⁵ (74.19) > Na³⁰ (70.96) > Cip⁵ (41.93) > Te³⁰ (38.70) = Do³⁰ (38.70) > Cot²⁵ (32.35) > Ak³⁰ (22.58) > C³⁰ (16.12) > Gen¹⁰ (12.90). In our study least resistance was observed against gentamycin, which is also in accordance with the previous study done by Silva group in Portugal [7]. In addition 10% resistance was found in the urban waste, Usha group in Indian hospital effluent was found to have 27.2% resistance [2,7]. Results showed and it can be assumed that such bacterial genera are more prone to other group

of antibiotics instead of aminoglycosides group. With the increasing use of antibiotics we came across the MAR index which showed us that how the irrational use of antibiotics is not only degrading the particular environment but also making the pathogenic microbes resistant to it. Regular monitoring would be useful in managing the increasing antibiotic resistance in localities to protect public health. Concerned authorities should put major measures to make public aware regarding downfalls of irrational consumption of antibiotics, to protect our environment and for the well-being of humanity.

<i>Enterobacteriaceae</i>	Resistance Pattern	MAR Index
<i>Escherichia coli</i>	AmpCfmCot	0.27
<i>Escherichia coli</i>	NaTeGenAkCfm	0.45
<i>Escherichia coli hermanii</i>	AmpNa	0.18
<i>Escherichia coli hermanii</i>	AmpCipNaTeDoCfm	0.54
<i>Escherichia coli, inactive</i>	AmpCipNaTeDoCazCfm	0.63
<i>Escherichia coli, inactive</i>	AmpCipNaCazCfm	0.45
<i>Escherichia coli, inactive</i>	AmpNa	0.18
<i>Escherichia coli, inactive</i>	AmpNaTeDoCazCot	0.54
<i>Escherichia coli, inactive</i>	AmpCipNaTeDoCazGenCfmC	0.81
<i>Escherichia coli, inactive</i>	AmpCipNaCazAkCfm	0.54
<i>Providencia rustigianii</i>	NaCazCfm	0.27
<i>Providencia rustigianii</i>	AmpNaCazCfmCot	0.45
<i>Providencia rustigianii</i>	AmpNaCazCfmCot	0.45
<i>Proteus mirabilis</i>	AmpCipNaCazAkCfmCot	0.63
<i>Proteus mirabilis</i>	AmpNaDoCazAkCfmCCot	0.72
<i>Proteus mirabilis</i>	AmpCipNaCazGenAkCfmC	0.72
<i>Proteus mirabilis</i>	AmpCazCfmCot	0.36
<i>Serratia plymuthica</i>	AmpNa	0.18
<i>Serratia plymuthica</i>	AmpCipNaCazCfm	0.45
<i>Proteus vulgaris</i>	AmpNaTeDoCazAk	0.54
<i>Proteus vulgaris</i>	AmpCipNaTeDoCazCfm	0.63
<i>Yersinia frederiksenii</i>	AmpCaz	0.18
<i>Yersinia frederiksenii</i>	AmpCipNaTeDoCfm	0.54
<i>Proteus penneri</i>	AmpNaTeDoCazCfmC	0.63
<i>Kluyvera ascorbata</i>	AmpCazCfmCot	0.36
<i>Budivicia aquatica</i>	AmpTeDoCazGenCfmCot	0.63
<i>Escherichia vulveris</i>	AmpCazCfm	0.27
<i>Salmonella choleraesi</i>	AmpCipCaz	0.27
<i>Yersinia pseudotuberculosis</i>	AmpCipCazAkCfmCCot	0.63
<i>Shigella sonnei</i>	AmpTeDoCazCfmCot	0.54
<i>Hafnia alvei</i>	AmpCipNaTeDo	0.45

[Table/Fig-1]: Multiple antibiotic resistance (MAR) index and resistance pattern of *Enterobacteriaceae* isolated from the raw sewage; AMP= Ampicillin; CFM= Cefixime; CAZ= Ceftazidime; C= Chloramphenicol; CIP= Ciprofloxacin; COT= Co-Trimoxazole; DO= Doxycycline Hydrochloride; GEN= Gentamicin; NA= Nalidixic acid; TE= Tetracycline

REFERENCES

- [1] Sieckmann DG, Reed ND, Georgi CE. Transferable drug resistance among *Enterobacteriaceae* isolated from human urinary tract infections. *Appl Microbiol* 1969;17(5):701-06.
- [2] Usha K, Kumar E, Gopal DS. Occurrence of various beta- lactamase producing Gram negative bacilli in the hospital effluent. *Asian J Pharm Clin Res*. 2013;6(3):42-46.
- [3] Central Pollution Control Board. Guide Manual: Water and wastewater analysis. Available from: http://www.cpcb.nic.in/upload/NewItems/NewItem_171_guidemannualw%26wwwanalysis.pdf >. Accessed 02 September 2014.
- [4] Bauer AW, Kirby WMM, Sherris JC, Turck M. Antibiotics susceptibility testing by a standardized single disk method. *Am J Clin Pathol*. 1966;45(4):493-96.
- [5] Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Second Informational Supplement. CLSI document M100-S21. Wayne, PA: 2012.
- [6] Krumperman PH. Multiple antibiotic resistance indexing of *Escherichia coli* to identify high- risk sources of fecal contamination of foods. *Appl Environ Microbiol*. 1983;46(1):165-70.
- [7] Silva MFD, Moreira IV, Pajuelo MG, Nunes OC, Manaia CM. Antimicrobial resistance patterns in *Enterobacteriaceae* isolated from an urban wastewater treatment plant. *FEMS Microbiol Ecol*. 2007;60:166-76.

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